



Department of Veterinary Pathobiology

DNA Technologies Core Lab

VMR Building 1197, Room 226

College Station, TX 77843-4467

<http://vetmed.tamu.edu/dnacore/bovid-dna-testing>

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Automated DNA Sequencing Request

Date : _____ Department: _____

Requested by: _____ Telephone: _____

Principal Investigator: _____ Email: _____

PO#: _____ Invoice #: _____

DNA Sequencing:

PLEASE NOTE: The quality and quantity of nucleotide sequence is determined by the template clean-up, quantitation and reaction parameters. Please provide sample information on the back of this form. Samples will be kept for 10 business days then discarded.

Sequencing – We run sequencing reaction.

Plasmid DNA 400-500ng. PCR products 100-150ng.

_____ RXNs@ 8.00= _____

Ready-To-Run – Client-preformed sequencing reaction

Note: remove excess dNTPs and dry sample thoroughly

_____ RXNs@ 2.00= _____

Consumables:

Dye Terminator Pre-Mix –

10 Reaction Kit (80uL)

_____ KITs@95.00= _____

100 Reaction Kit (800uL)

_____ KITs@ 925.00= _____

Sub-total= _____

Make checks payable to:

Texas Agrilife Research

Veterinary Pathobiology

4467 – TAMU

College Station, TX 77843-4467

Total Charges \$ _____

Sequencing Sample Information

Please fill out **ALL** columns using concentration units as specified – see recommended protocol below for template and primer concentrations required to perform your reactions

Sample Name	Template Name Type Conc. (ng/ μ l) (PCR Size)	Primer Name, Conc, T _m

Recommended Protocol for 10 μ L Sequencing Reaction

Reagent	Working Concentration	Volume
Big Dye		4.0 μl (or use 2.0 μ L Big Dye + 2.0 μ L $\frac{1}{2}$ Big Dye)
Template Single-Stranded DNA Double-Stranded DNA PCR Product DNA	50-100ng/ μ l 400-500ng/ μ l 10ng/ μ l per 100bp	1.0 μL (adjust up or down if template varies from listed optimal concentrations)
Primer	5-10 μ M	1.0 μL (adjust up or down if primer varies from listed optimal concentration)
Deionized Water (may substitute ABI sequencing Buffer)		q.s.
Total Reaction Volume		10 μ l